

Setting Up the PTC Hood Workspace

Please outfit the hood in this order:

1. Set the file folder supports so the frame faces forward. If you set it up correctly, one long bar will be on the table in front of you. Tell students to consider this the line of aseptic handling. NOTHING open should go outside this line.
2. Drape the plastic over the top so the fold overhangs the upper bar by about 3-4 inches, forming a "sneeze-protector"
3. Spray the inside of the hood "floor, sides and ceiling, with 70% rubbing alcohol straight from the bottle.
4. Put a large waste beaker (has 3 protrusions) in the back. Students will pour the water rinse waste into the beaker IN the hood.
5. Put an erlenmeyer flask in the hood.
6. Put an empty glass 20X150 culture tube in the flask open end up.
7. Put a scalpel in the tube in the flask
8. Next put in a curved forceps
9. Last put in a large forceps
10. Fill the tube to the brim with the rubbing alcohol. This will mean that the scalpel and curved forceps are fully immersed in alcohol.
11. Place in the hood a 50 ml tube w/ small piece of styrofoam to hold it. Use the tube to dispense sterile water into the LARGE Petri Plate. One plate per team of 3 ONLY.
12. Put in a sterile water bottle. Spray the cap rim with some alcohol. Loosen the cap aseptically and WARN students to put the cap bottoms up in back of the hood when they get ready to dispense the water.
13. Spray everything again.
14. REMIND students that the hood is NOT sterile, just relatively aseptic. They must do their cutting on the INSIDE surface of the Petri plate. They can use the rim to set the instruments down to drain.
15. Each team will bring in their 3 culture tubes. The caps should be placed **lid up** like in this Petri plate sketch.

